



Institut für Medizinische Mikrobiologie, Virologie und Hygiene



BS-Experten Netzwerktreffen und refresher, Kassel 10.11.2023

Was bringt uns die syndromische PCR-Diagnostik?

Prof. Dr. med. Holger Rohde

Leitender Oberarzt

Leiter Arbeitsgruppe Staphylokokkeninfektionen

Director ESCMID AMR Action Committee

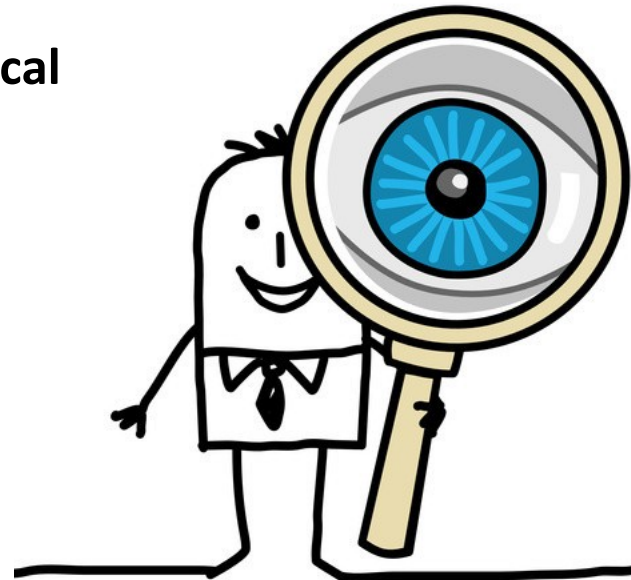
ESCMID Fellow



Universitätsklinikum
Hamburg-Eppendorf

Das technische Auge der modernen Mikrobiologie hat sich grundlegend verändert:

- Massenspektrometrie (MALDI-TOF)
- **Molekulare Assays (Hochdurchsatz, syndromische Panels)**
- Beschleunigte Empfindlichkeitsprüfung (RAST)
- Next generation sequencing (WGS, clinical metagenomics)



Chances	Threats
Schneller Ergebnisse (klinische Signifikanz)	
Reduktion falsch- negativer Befunde	
Gezielte Therapie	

Chances	Threats
Schneller Ergebnisse (klinische Signifikanz)	Hohe Kosten
Reduktion falsch- negativer Befunde	Falsch-positive / Übertherapie
Gezielte Therapie	Nicht indizierte AB Therapien

Bei der Durchführung syndromorientierter, molekularer Analytik ist die Definition eines “sweet spots” zur Erreichung optimaler Effekte notwendig.

Ein erheblicher Teil der Gelenkinfektionen bleibt ätiologisch ungeklärt (d. h. kulturnegativ).

Infektionstyp	% Kultur-negativ	Referenz
Prosthetic joint infection	5 - 34	Tande and Patel, CMR 2014
Native joint septic arthritis	15 - 60	Richebé et al, BMJ 2022 McBride et al., CID 2020

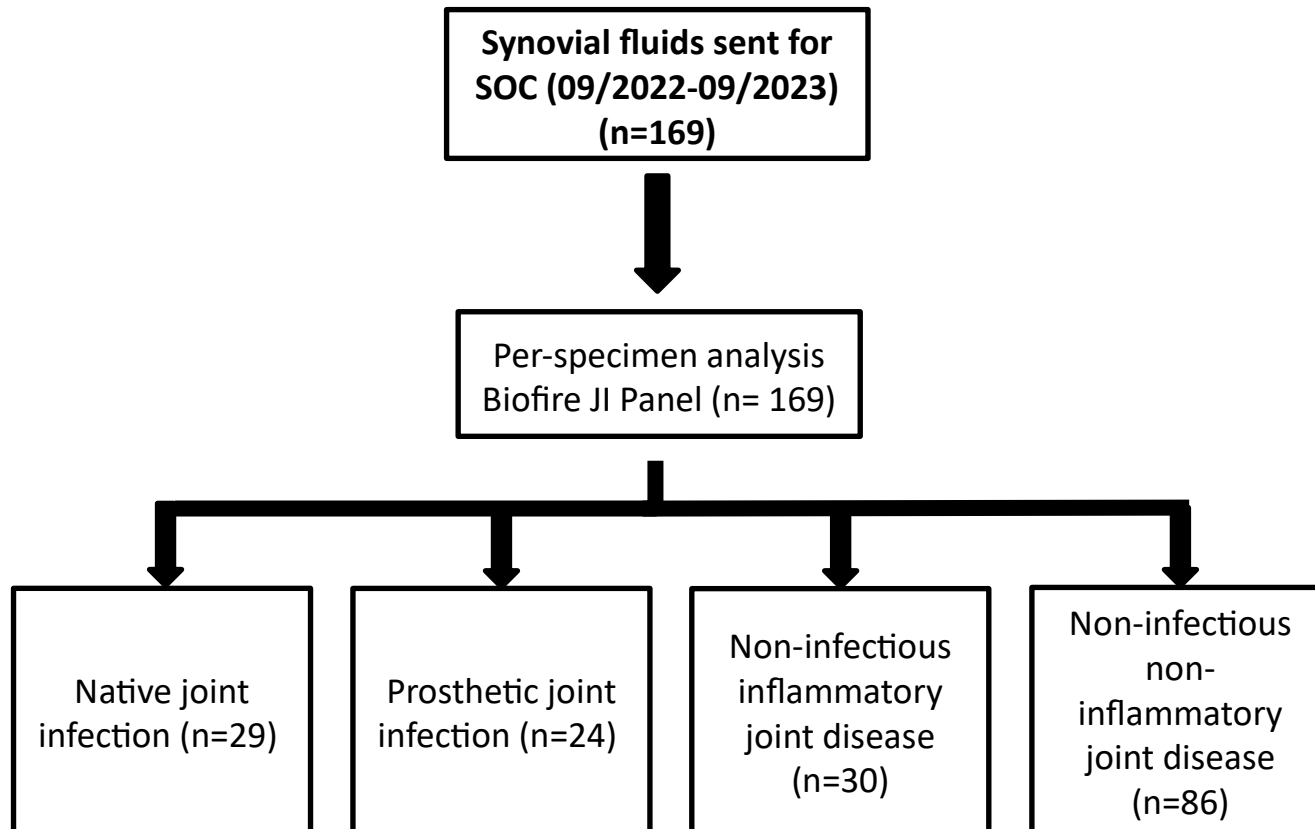
Table 1

Overview of pathogens identified in BJI specimens using SOC or molecular techniques.

Species	Pathogens detected ^a		
	SOC	mPCR	16S rDNA PCR
Monomicrobial infections (n=38)			
<i>S. aureus</i>	11	14	6
CoNS	7	4	2
<i>C. acnes</i>	1	6	0
Enterobacterales (total)	9	8	3
<i>Serratia marcescens</i>	3	2	0
<i>Enterobacter cloacae</i>	4	4	2
<i>Klebsiella pneumoniae</i>	2	2	1
<i>S. dysgalactiae</i>	1	1	1
<i>S. agalactiae</i>	3	3	2
Other (total)	6	1	0
<i>E. faecalis</i>	5	5	1
<i>C. subterminale</i>	1	0	0
Polymicrobial infections (n=2)	2	1	1
<i>S. aureus</i>	1	1	1
<i>P. mirabilis</i>	1	1	1
<i>S. epidermidis</i>	1	0	0
<i>S. haemolyticus</i>	1	0	0

^a In total, 126 specimens were analyzed by each method.

Bedeutung eines spPCR Panels bei der Diagnostik von Gelenkinfektionen: eine prospektive Kohortenstudie



SOC positive (%) n=12 (41)

n=15 (62)

n=0

n=0

JI positive (%) n=16 (55)

n=20 (83)

n=0

n=1

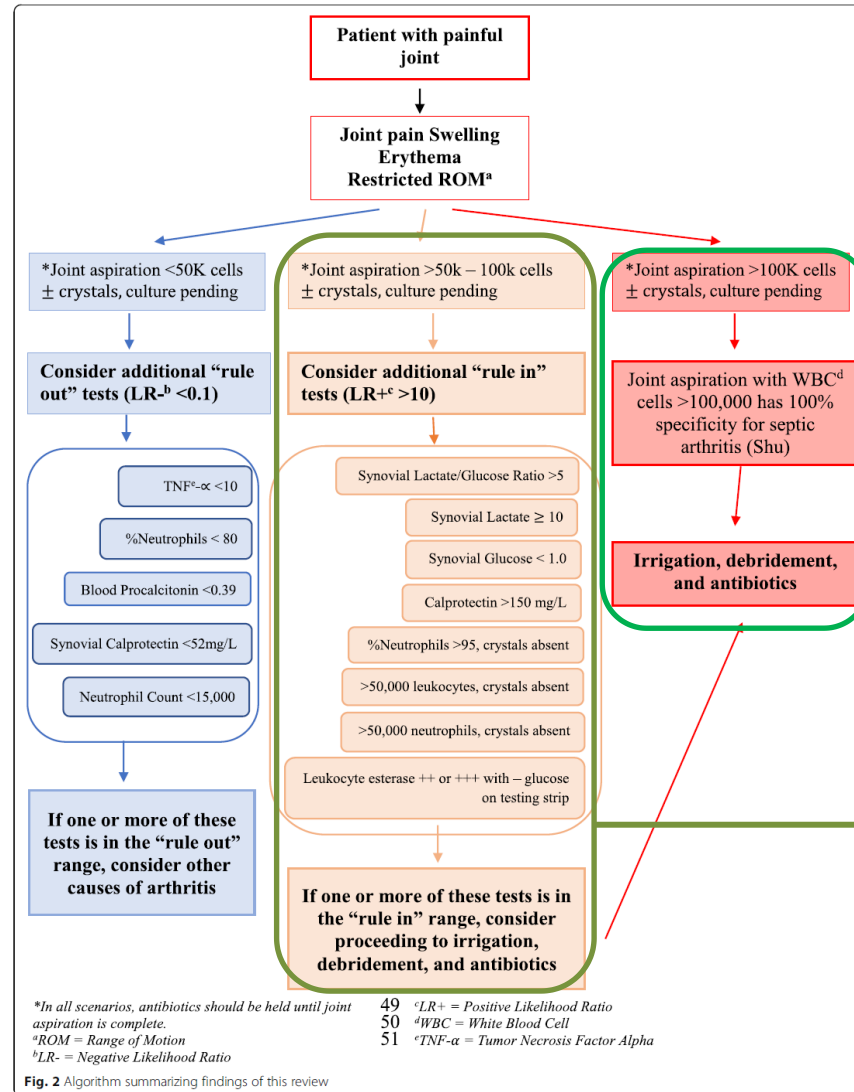


Fig. 2 Algorithm summarizing findings of this review

spPCR als Zusatz zur schnellen und sensitiven Analytik.

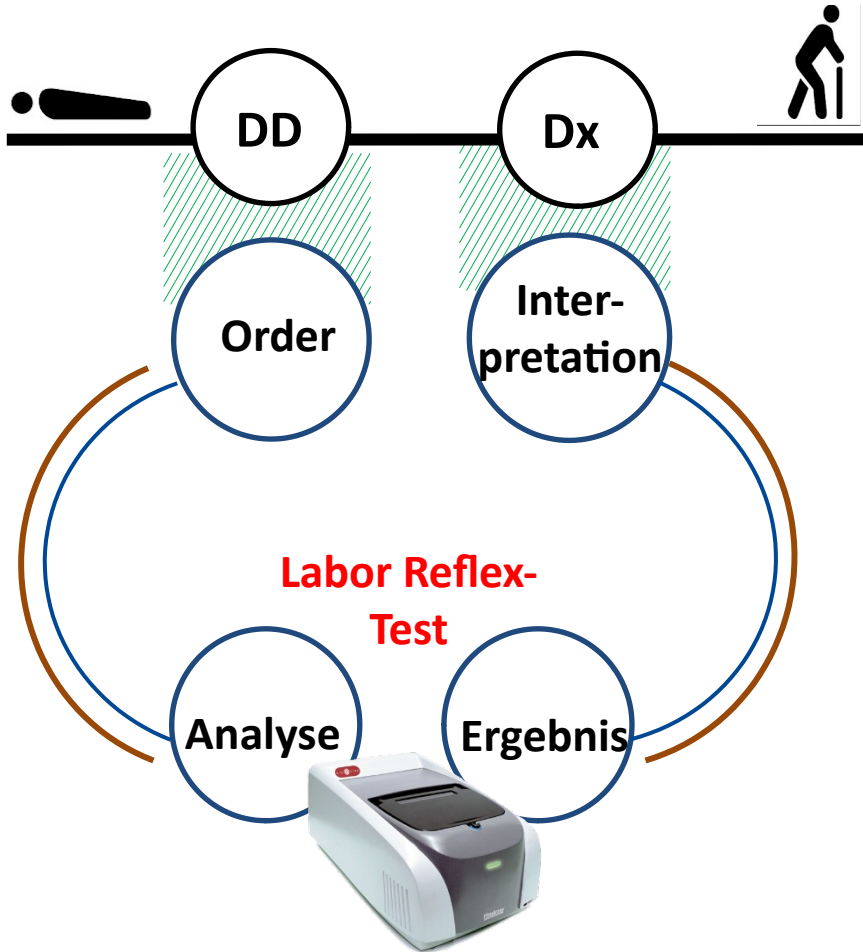
spPCR als „rule in“ Instrument.

	Number of colonies		
	<5	5–50	>50
	Available results, <i>n</i> = 94		
Culture and mPCR positive concordant results <i>n</i> = 102 (%)	13 (13.8)	32 (34.0)	49 (52.2)
	Available results, <i>n</i> = 65		
Culture positive and mPCR negative results <i>n</i> = 115 (%)	30 (39.4)	21 (37.9)	14 (22.7)

“.....it is essential that local implementation of the (PCR) panels is done in partnership with clinicians to ensure that there is a clear understanding of test characteristics, result interpretation, and appropriate test utilization.”

Hanson, JCM 2016

	Number of colonies		
	<5	5–50	>50
Available results, <i>n</i> = 94			
Culture and mPCR positive concordant results <i>n</i> = 102 (%)	13 (13.8)	32 (34.0)	49 (52.2)
Available results, <i>n</i> = 65			
Culture positive and mPCR negative results <i>n</i> = 115 (%)	30 (39.4)	21 (37.9)	14 (22.7)



	UKE-PCR	Biofire	Kultur
<i>E. coli</i>	0	0	0
<i>H. Influenzae</i>	0	0	0
<i>L. monozytogenes</i>	1	1	1
<i>N. meningitidis</i>	3	3	0
<i>S. agalactiae</i>	2	2	0
<i>S. pneumoniae</i>	12	13	6
CMV	0	0	
Enteroviren	6	5	
HSV1	0	0	
HSV2	2	2	
HHV6	3	3	
Parechovirus	0	0	
VZV	3	3	
<i>C. neoformans</i>	1	1	1

18 Monate: 4623 Liquorproben



Gramfärbung:
Liquorpleozytose?

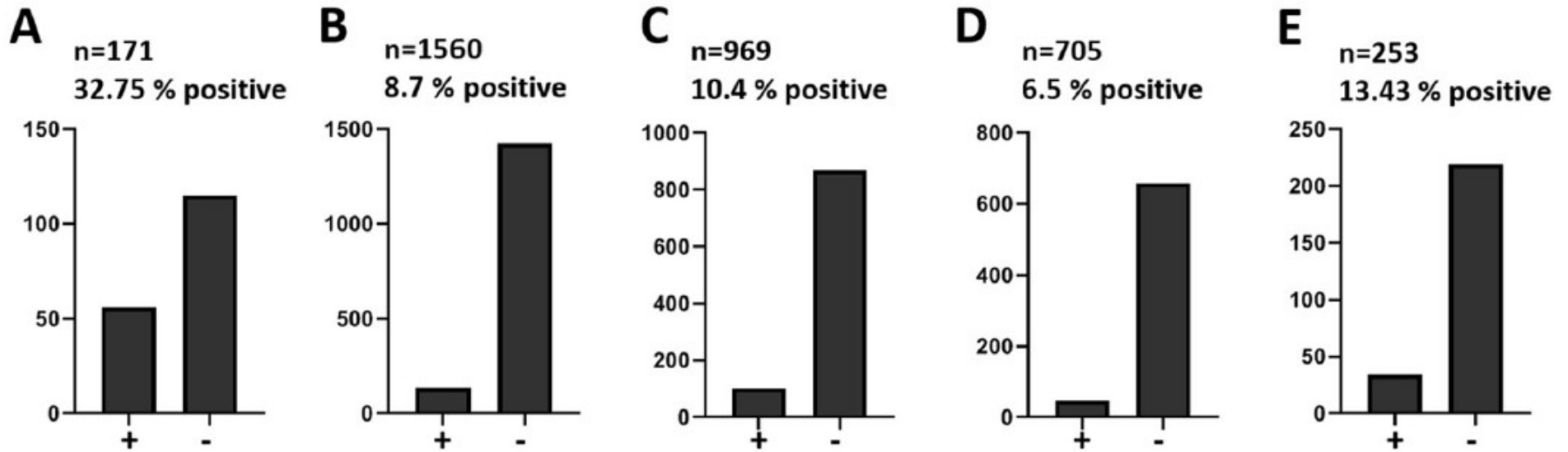


**Biofire
Filmarray**



**Standard-
diagnostik (Kultur)**

n=171 Proben analysiert

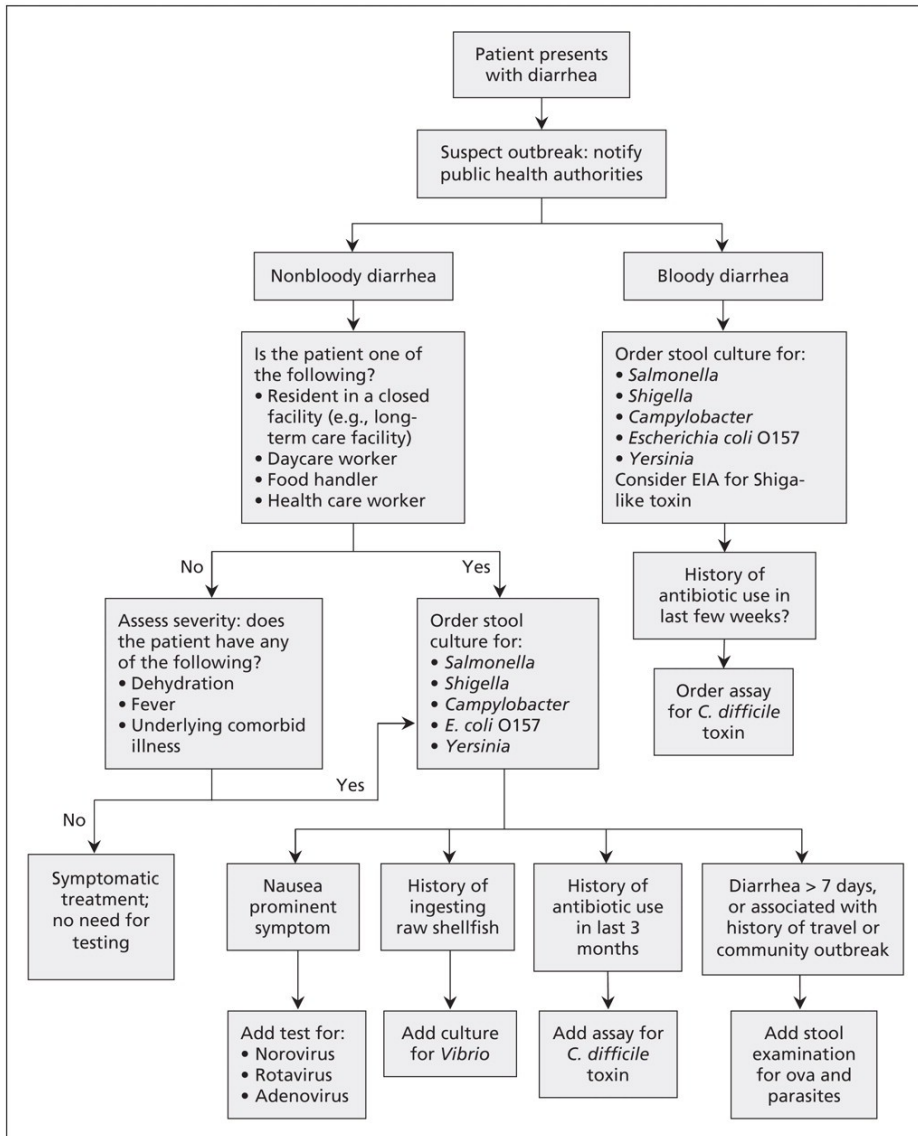


Pfefferle, Rohde et al., BMC Infect Dis 2020

- “...the per-patient yield of the ME panel increased 61.8% from an estimated 11.5% (53/459) positivity rate with no testing restriction to 18.6% (49/263) with criteria in place ($P < 0.01$).”

Broadhurst et al., JCM 2020

Risikoprofil getriebener Algorithmus zum zielgerichteten Einsatz konventioneller Methoden: Anforderer



Für den optimalen Einsatz empfehlen klinische Guidelines selektive Testungen bei GI-Infektionen: à la carte Strategie.

Hatchette und Farina CMAJ 2011

In vielen klinischen Kontexten ist die Festlegung der Untersuchungsmethode/ des Untersuchungsumfangs traditionell auf Seite des Klinikers verankert.

Table 2. Unrequisitioned bacteria and parasites detected by xTAG GPP.

Target	No. not requested by physician	% of total additional positives
<i>Salmonella</i>	6	46
<i>Campylobacter</i>	10	40
<i>Shigella</i>	4	24
<i>Clostridium difficile</i> Toxin A/B ^a	32	74
ETEC	12	63
<i>E. coli</i> O157	6	55
STEC	13	81
<i>Yersinia enterocolitica</i>	1	100
<i>Giardia</i>	31	79
<i>Entamoeba histolytica</i>	10	71
<i>Cryptosporidium</i>	12	100
Overall	137	65

➤ **Untersuchungsumfang und Testcharakteristika häufig unbekannt – dies führt zu fehlerhaften / unvollständigen Anforderungen.**

^a18/32 positive specimens collected from subjects <3 years old.

Syndromische multiplex PCR für den Nachweis von gastrointestinalen Pathogenen: one-stop-shop

TABLE 4 FDA-approved/cleared multiplex gastrointestinal panels^a

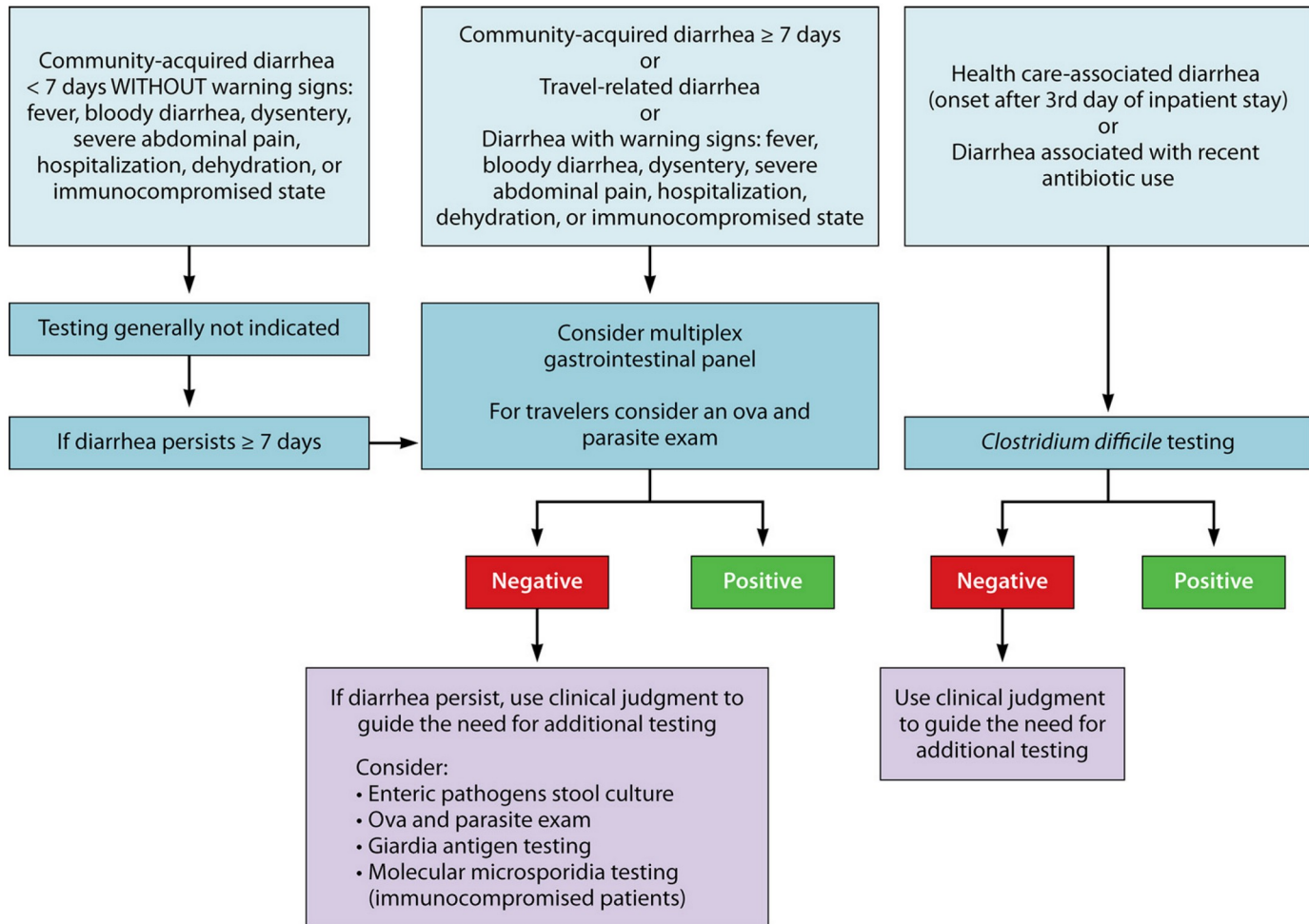
Parameter	Verigene EP	Luminex GPP	BioFire GIP
Analysis platform	Verigene system	Magpix or Luminex 100/200 system	FilmArray system or FilmArray Torch
Acceptable specimen type	Stool in Cary-Blair medium	Fresh stool or stool in Cary-Blair medium	Stool in Cary-Blair medium
No. of targets	9	14	22
Ability to detect pathogen			
Bacteria			
<i>Campylobacter</i> species	✓	✓	✓
<i>Salmonella</i> species	✓	✓	✓
<i>Shigella</i> species/enteroinvasive <i>E. coli</i> ^b	✓	✓	✓
<i>Vibrio</i> species	✓		✓
<i>Vibrio cholerae</i>		✓	✓
<i>Yersinia enterocolitica</i>	✓		✓
<i>Escherichia coli</i> O157		✓	✓
Enterotoxigenic <i>E. coli</i>		✓	✓
Enteropathogenic <i>E. coli</i>			✓
Enteraggregative <i>E. coli</i>			✓
<i>Plesiomonas shigelloides</i>			✓
Shiga toxin-producing <i>E. coli</i> (stx ₁ -stx ₂)	✓ ^c	✓	✓
<i>Clostridium difficile</i> (toxin A/B)		✓	✓
Viruses			
Norovirus GI/GII	✓	✓	✓
Rotavirus A	✓	✓	✓
Astrovirus			✓
Adenovirus 40/41		✓	✓
Sapovirus			✓
Parasites			
<i>Cryptosporidium</i> species		✓	✓
<i>Entamoeba histolytica</i>		✓	✓
<i>Giardia lamblia</i>		✓	✓
<i>Cyclospora cayetanensis</i>			✓
No. of samples (throughput)	1–32 (scalable)	24	1–12 (scalable)
Time to result (h)	<2	~5	~1

^aEP, enteric pathogens; GPP, gastrointestinal pathogen panel; GIP, gastrointestinal panel.

^bThe Verigene EP and Luminex GPP do not specifically target enteroinvasive *E. coli*.

^cThe Verigene EP has separate targets for stx₁ and stx₂.

Syndromische multiplex PCR: vereinfachte Anforderung, transparenteres Output



1. This algorithm should not be used for chronic diarrhea (duration >30 days).
2. For ova and parasite exams, submit 3 stool samples collected on separate days for maximum sensitivity.
3. During the summer, consider molecular detection of Shiga toxin in fecal samples for children with diarrhea even if they do not have bloody diarrhea, are not toxic-appearing, and diarrhea has been present <7 days.

Clinical impact of syndromic molecular point-of-care testing for gastrointestinal pathogens in adults hospitalised with suspected gastroenteritis (GastroPOC): a pragmatic, open-label, randomised controlled trial



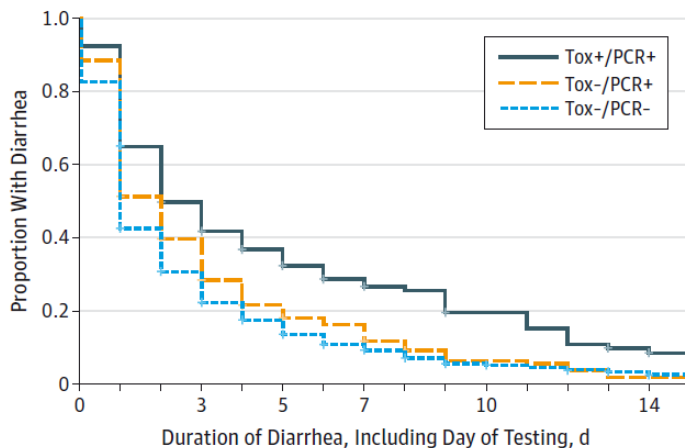
Nathan J Brendish, Kate R Beard, Ahalya K Malachira, Alex R Tanner, Langizya Sanga-Nyirongo, Markus Gwiggner, J R Fraser Cummings, Helen E Moses, Tristan W Clark



- Single center, UK, Notaufnahme oder stationäre Patienten innerhalb 48h nach Aufnahme**
- 278 Patienten (138 Pat. FilmArray Gastrointestinal Panel (BioFire), 140 Pat. Kontrolle)**
- Time to result: (BioFire vs. Kont.) (median (IQR)): 1.7 h (1.5–2.0) vs. 44.7 h (21.2 – 66.1), sig**
- Pat. ohne Erregernachweis (55% in BioFire vs. 74% in Kontrolle):**
- Entisolation (PCR vs. Kont.): 73% vs. 34%, $p < 0.0001$
 - Zeit bis Entisolation: 0.6 Tage (0.3 – 1.8) vs. 2.2 Tage (1.2 – 3.2), $p < 0.0001$
- Antibiotikatherapie (BioFire vs. Kont.): 65% vs. 47%, $p = 0.0028$**

- Wahrscheinlichkeit des Nachweises von kausalen Pathogenen steigt im Vergleich zur *à la carte* Strategie.
 - Ergebnisse sind schneller verfügbar.
 - Assays können Einfluss auf das klinische Management nehmen (z.B. Isolationsmaßnahmen).
- Zusätzliche Kosten.
 - Klinische Signifikanz von Ergebnissen unklar (Nachweis von > 1 Pathogen [z.B. zusätzliche EPEC Nachweise in 10 – 30 %]).
 - Risiko falsch-positiver Befunde.

Polage et al., JAMA Int Med 2015

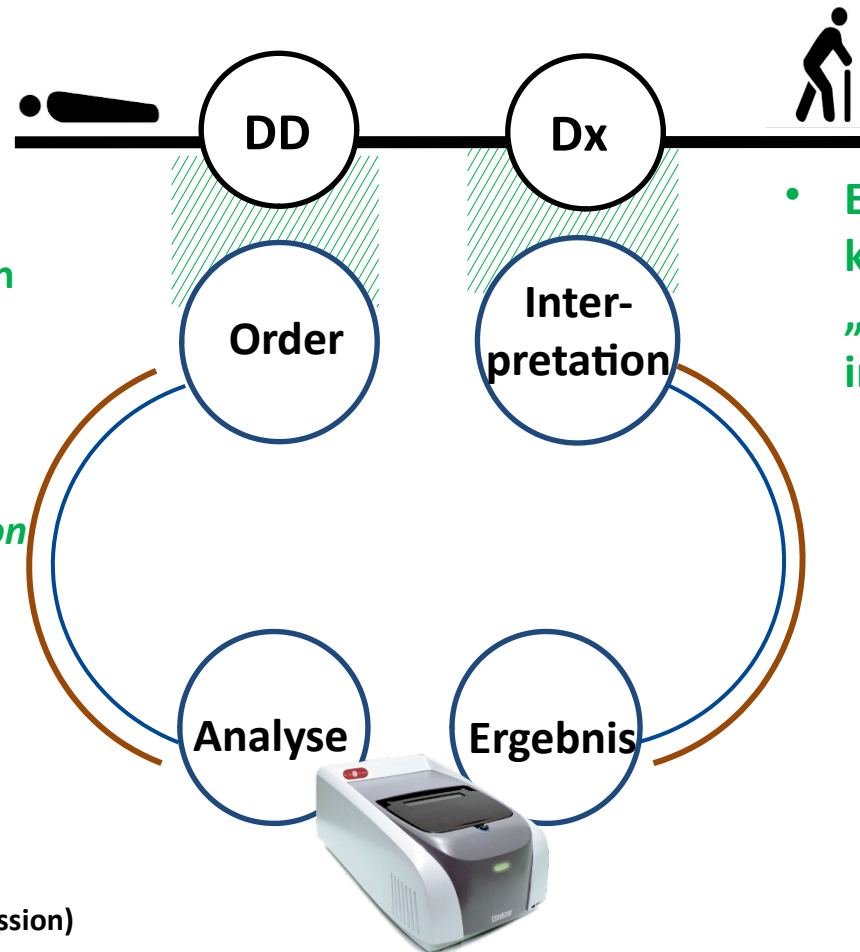


No. at risk							
Tox+/PCR+	131	62	41	29	25	8	
Tox-/PCR+	162	60	29	21	10	2	
Tox-/PCR-	1123	328	172	99	42	23	

Der breite Einsatz molekularer (multiplex) Diagnostik kann zu einer Erhöhung falsch-positiver Befunde führen: Übertherapie droht.

Messacar et al., JCM 2017

PCR (Panel) Diagnostik: so einfach?



- Ergebnisintegration in klinische Handlungsfade: „Intention to test is intention to act“.

- Indikationsstellung: Verbindliche Festlegung diagnostischer Algorithmen (SOP).
- Kontinuierliche Überwachung des Testeinsatzes und der Performance (*Lab utilization committee*)

TABLE 3 FDA-approved/cleared multiplex respiratory panels^a

Parameter	FilmArray	Verigene	x-TAG RVP	x-TAG RVP Fast	NxTAG-RPP	eSensor RVP	ePlex
Analysis platform	FilmArray system or FilmArray Torch	Verigene system	Luminex 100/200	Luminex 100/200	Luminex Magpix	eSensor	ePlex system
No. of targets	20	16	12	8	20	14	17
Ability to detect pathogen							
Virale Erreger	Viruses						
	Adenovirus	✓	✓	✓	✓	✓ (differentiates subgroup B/E from C)	✓
	Coronavirus						✓
	Coronavirus HKU1	✓				✓	
	Coronavirus NL63	✓				✓	
	Coronavirus 229E	✓				✓	
	Coronavirus OC43	✓				✓	
	Human bocavirus					✓	
	Human metapneumovirus	✓	✓	✓	✓	✓	✓
	Influenza A virus	✓	✓	✓	✓	✓	✓
	Subtype H1	✓	✓	✓	✓	✓	✓
	Subtype H3	✓	✓	✓	✓	✓	✓
	Subtype 2009 H1N1	✓	✓	✓	✓	✓	✓
	Influenza B virus	✓	✓	✓	✓	✓	✓
	Parainfluenza virus 1	✓	✓	✓	✓	✓	✓
	Parainfluenza virus 2	✓	✓	✓	✓	✓	✓
	Parainfluenza virus 3	✓	✓	✓	✓	✓	✓
	Parainfluenza virus 4	✓	✓	✓	✓	✓	✓
	Respiratory syncytial virus	✓			✓		
	Respiratory syncytial virus A		✓	✓		✓	✓
Respiratory syncytial virus B		✓	✓		✓	✓	
Rhinovirus/enterovirus	✓	✓	✓	✓	✓	✓	
Bakterien	Bacteria						
	<i>Chlamydomphila pneumoniae</i>	✓			✓		✓
	<i>Mycoplasma pneumoniae</i>	✓			✓		✓
	<i>Bordetella pertussis</i>		✓				
	<i>Bordetella parapertussis-Bordetella bronchiseptica</i>		✓				
	<i>Bordetella holmesii</i>		✓				
Time to result (h)	~1	~2-3	~8	~6	~4	~6	~1.5

^aThe acceptable specimen type for all panels is a nasopharyngeal swab. RVP, respiratory virus panel; RPP, respiratory pathogen panel.

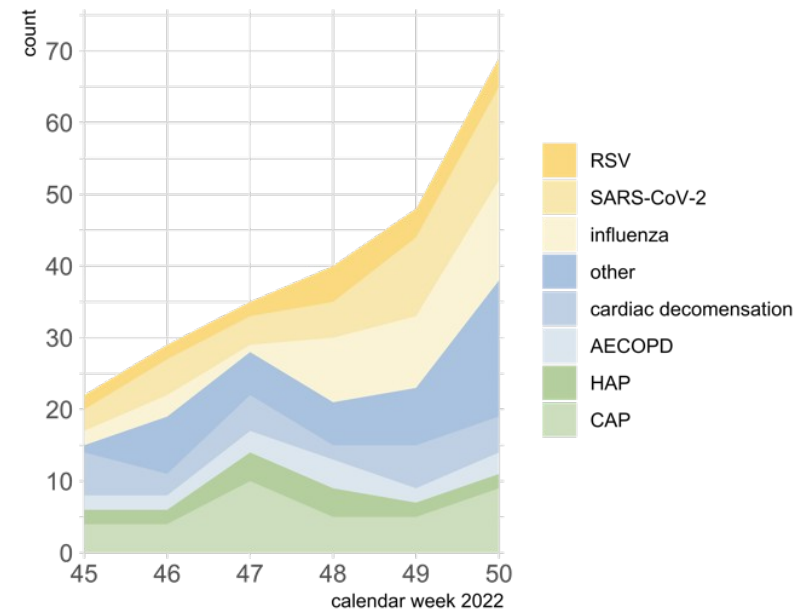
Empfehlungen zur Durchführung mikrobiologischer Analytik bei der CAP

Community-acquired pneumonia: Initial evaluation and site of care based on severity assessment

	Severity score*	Site of care	Microbiologic evaluation
Mild	PSI: I or II or CURB-65: 0 [¶]	Ambulatory care	<ul style="list-style-type: none"> COVID-19 testing during the pandemic Influenza testing (when incidence is high and results would change management)^Δ Otherwise, testing is usually not needed
Moderate	PSI: III or IV or CURB-65: 1 [¶] to 2	General medical ward	<ul style="list-style-type: none"> Blood cultures Sputum Gram stain and culture Urine streptococcal antigen <i>Legionella</i> testing[◊] Respiratory viral panel during respiratory virus season[§] COVID-19 testing[‡] HIV screening[‡]
Severe	PSI: IV or V or CURB-65: ≥3 and/or Fulfillment of ATS/IDSA criteria for ICU admission [†]	ICU	<ul style="list-style-type: none"> Blood cultures Sputum Gram stain and culture Urine streptococcal antigen test <i>Legionella</i> testing[◊] Respiratory viral panel[§] Bronchoscopy specimens for Gram stain, fungal stain, aerobic, fungal culture, and molecular testing (when feasible)^{**} COVID-19 testing[‡] HIV screening[‡]

Klompas, UpToDate 2021

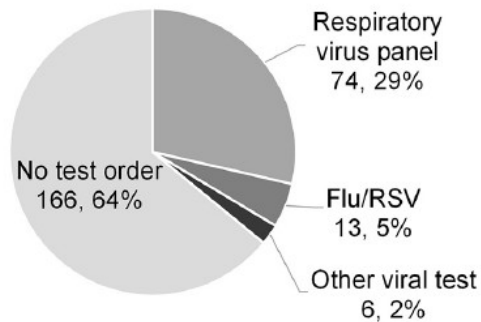
Ungewöhnliche Epidemiologie viraler CAP Erreger Ende 2022



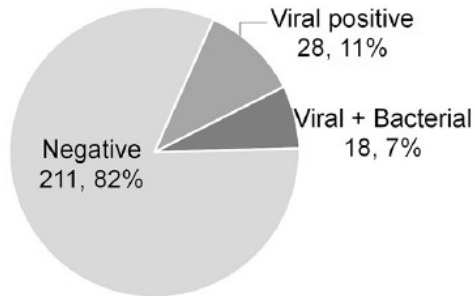
Koch, Rohde, Lütgehetmann, in revision 2023

Verbesserte diagnostische Ausbeute

A Specimens with SOC Order for Viral Target

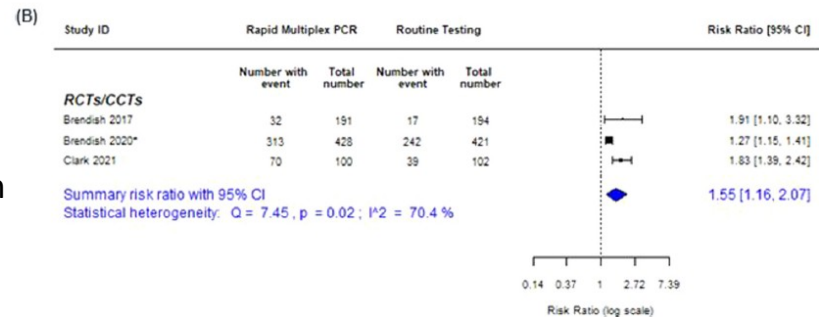
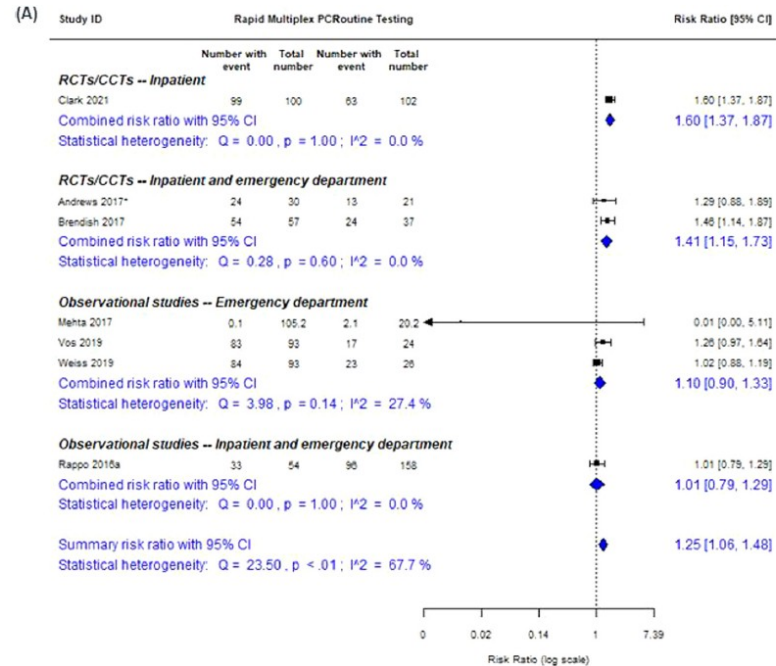


B Specimens with a Viral Target Detected



11/46 Virus-positiven Proben wären durch konventionelle Anforderungsstrategien identifiziert worden.

Management Konsequenzen



THE LANCET Respiratory Medicine

Volume 10, Issue 9, September 2022, Pages 877-887

Articles

Fast multiplex bacterial PCR of bronchoalveolar lavage for antibiotic stewardship in hospitalised patients with pneumonia at risk of Gram-negative bacterial infection (Flagship II): a multicentre, randomised controlled trial

Andrei M Darie MD,^a Prof Nina Khanna MD,^{b,c} Kathleen Jahn MD,^a Michael Osthoff MD,^d Prof Stefano Bassetti MD,^d Mirjam Osthoff MD,^a Desiree M Schumann PhD,^a Prof Werner C Albrich MD,^f Prof Hans Hirsch MD,^e Prof Martin Brutsche MD,^g Leticia Grize PhD,^a Prof Michael Tamm MD,^a Prof Daiana Stolz MD,^{a,c,h}  



Contents lists available at [ScienceDirect](https://www.sciencedirect.com)

Journal of Infection

journal homepage: www.elsevier.com/locate/jinf

Molecular point-of-care testing for lower respiratory tract pathogens improves safe antibiotic de-escalation in patients with pneumonia in the ICU: Results of a randomised controlled trial

Stephen Poole^{a,b,*}, Alex R Tanner^b, Vasanth V Naidu^b, Florina Borca^{a,c}, Hang Phan^c, Kordo Saeed^{b,d}, Michael P W Grocott^{a,d,e}, Ahilanandan Dushianthan^{a,d,e}, Helen Moyses^a, Tristan W Clark^{a,b,d,f}

^aNIHR Southampton Biomedical Research Centre, University Hospital Southampton NHS Foundation Trust, Southampton, United Kingdom

^bDepartment of Infection, University Hospital Southampton NHS Foundation Trust, Southampton, United Kingdom

^cClinical Informatics Research Unit, University of Southampton, Southampton, United Kingdom

^dSchool of Clinical and Experimental Sciences, Faculty of Medicine, University of Southampton, Southampton, United Kingdom

^eGeneral Intensive Care Unit, University Hospital Southampton NHS Foundation Trust, Southampton, United Kingdom

^fNIHR Post-Doctoral Fellowship Programme, United Kingdom

- St Gallen, RCT, CAP, **208 Pat**
- Unyvero Hospitalized Pneumonia Panel + Therapieempfehlung
- **Inadäquate ABX (Prim. EP): 47h vs. 86h, sig**
- kein Unterschied Nebenwirkungen, Outcome

- UK Southampton, RCT, **200 ICU Pat** (68 Pat CAP)
- FilmArray Pneumonia *plus* Panel(BioFire), Mo-Fr Kernarbeitszeit (Befundergebnis: 1.7h vs. 66.7h)
- **Erregernachweis (PCR vs Kont): 71% vs 51%, sig**
- **Deeskalation (PCR vs Kont): 42% vs. 8%, sig**
- kein Unterschied Nebenwirkungen, Outcome

Panelanalytik ist zielführend wenn (messbare) Ziele im klinischen Management definiert worden sind!

The right test

The right patient

The right time

**Ist der Test für das
klinische Szenario
geeignet?**

**Wird die klinische
Versorgung des
Patienten durch das
Testergebnis
beeinflusst?**

**Wird das Ergebnis rechtzeitig
zur Verfügung stehen, um die
Behandlung optimal zu
beeinflussen?**

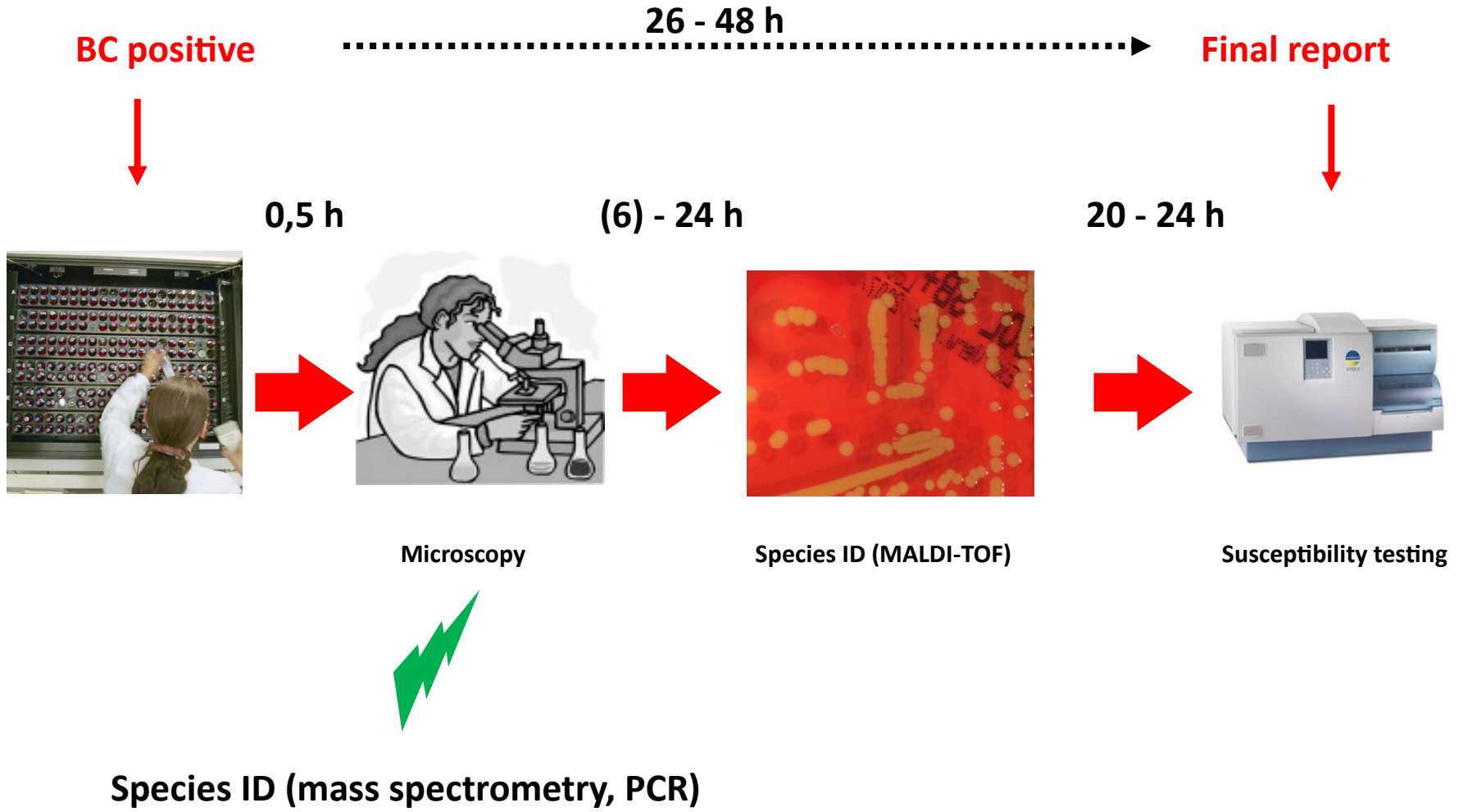
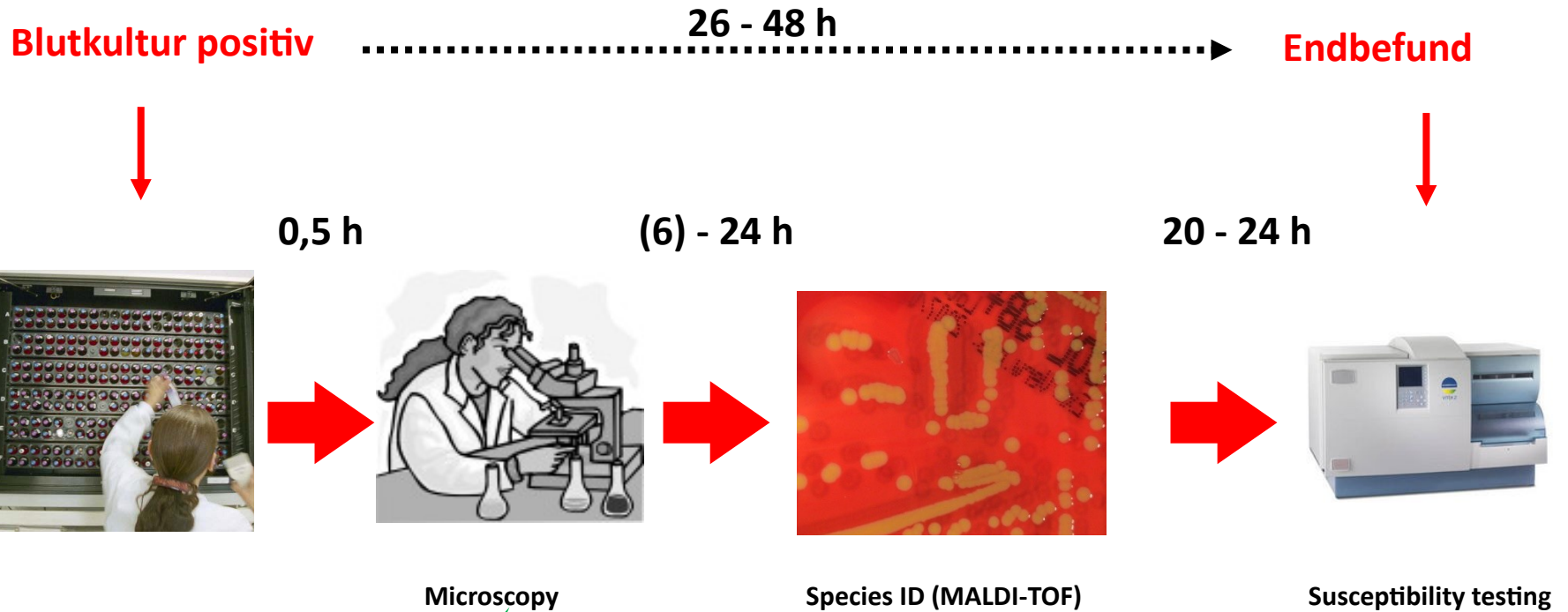


Table 1 Targets included in the BCID2 Panel

Gram negatives	Gram positives	Yeast	Antimicrobial resistance markers
<i>A. calcoaceticus-baumannii</i> complex	<i>Staphylococcus spp.</i>	<i>Candida albicans</i>	<i>mecA/C</i>
<i>Bacteroides fragilis</i>	<i>Staphylococcus aureus</i>	<i>Candida auris</i>	<i>mecA/C and MREJ (MRSA)</i>
<i>Haemophilus influenzae</i>	<i>Staphylococcus epidermidis</i>	<i>Candida glabrata</i>	<i>van A/B</i>
<i>Neisseria meningitidis</i>	<i>Staphylococcus lugdunensis</i>	<i>Candida krusei</i>	<i>blaCTX-M</i>
<i>Pseudomonas aeruginosa</i>	<i>Streptococcus spp.</i>	<i>Candida parapsilosis</i>	<i>blaKPC</i>
<i>Stenotrophomonas maltophilia</i>	<i>Streptococcus agalactiae</i>	<i>Candida tropicalis</i>	<i>blaIMP</i>
<i>Enterobacterales spp.</i>	<i>Streptococcus pyogenes</i>	<i>Cryptococcus neoformans/gattii</i>	<i>blaOXA-48</i>
<i>Enterobacter cloacae</i> complex	<i>Streptococcus pneumoniae</i>		<i>blaNDM</i>
<i>Escherichia coli</i>	<i>Enterococcus faecalis</i>		<i>blaVIM</i>
<i>Klebsiella aerogenes</i>	<i>Enterococcus faecium</i>		<i>mcr-1</i>
<i>Klebsiella oxytoca</i>	<i>Listeria monocytogenes</i>		
<i>Klebsiella pneumoniae</i> group			
<i>Proteus spp.</i>			
<i>Salmonella</i>			
<i>S. marcescens</i>			

MRSA = Methicillin Resistant *Staphylococcus aureus*



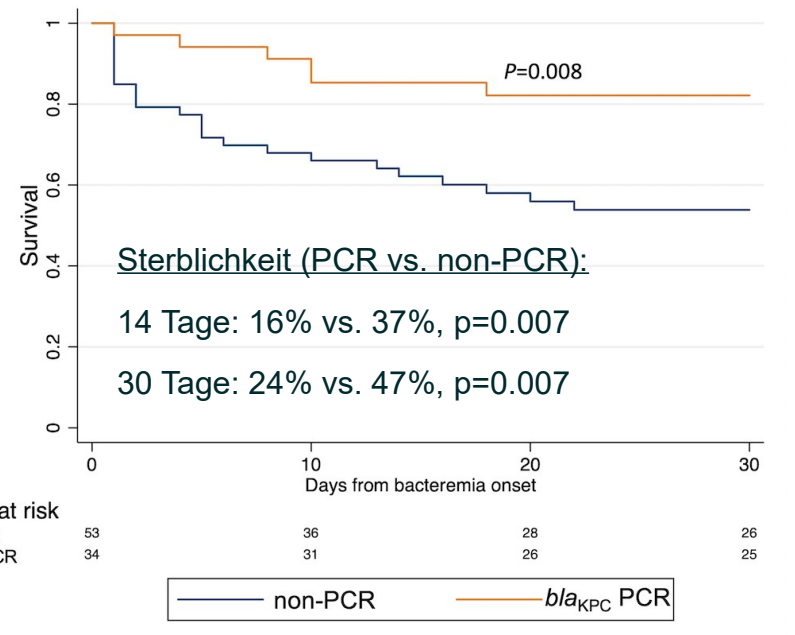
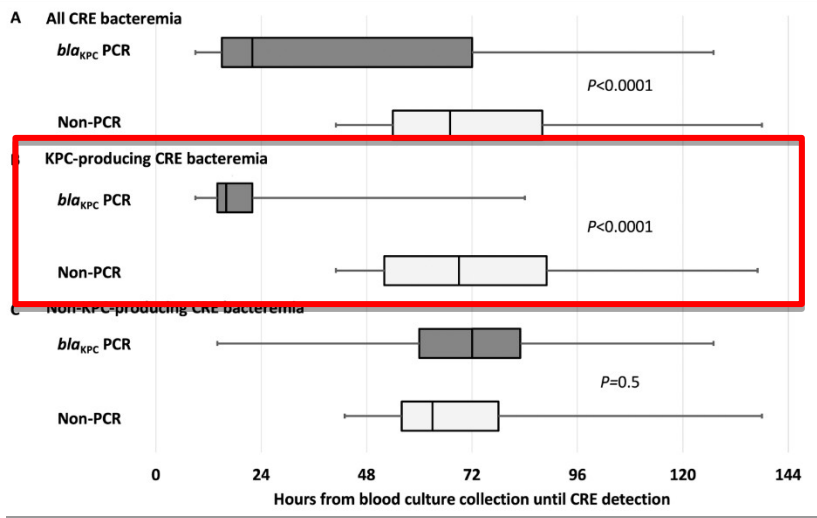
**Antibiotika
therapie**

Molecular test for species ID +
susceptibility prediction

Optimal

- Beobachtungsstudie, 8 Kliniken New York & New Jersey, 2016-2018
- In 3 Kliniken bla_{KPC} PCR (BCID, BioFire) etabliert
- 137 Pat mit CRE-BSI (davon 89 Pat. KPC+)
 - 51 Pat. haben bla_{KPC} PCR bekommen
 - 32 Pat KPC positiv getestet (1x falsch neg)

Schnellere Befundmitteilung: 16h vs. 64h



7.30 a.m. – 1 p.m.

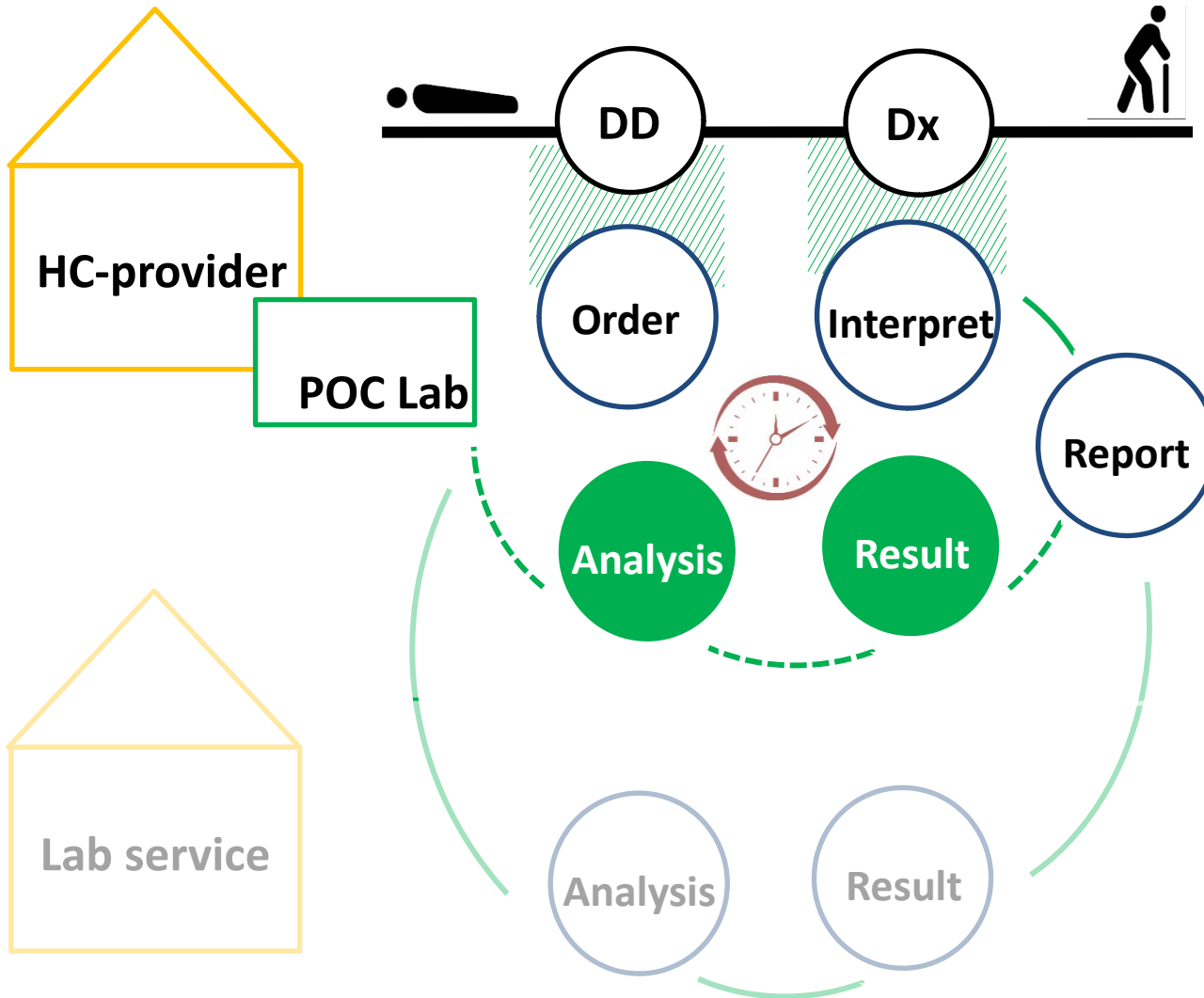
Same day: species ID + susceptibility

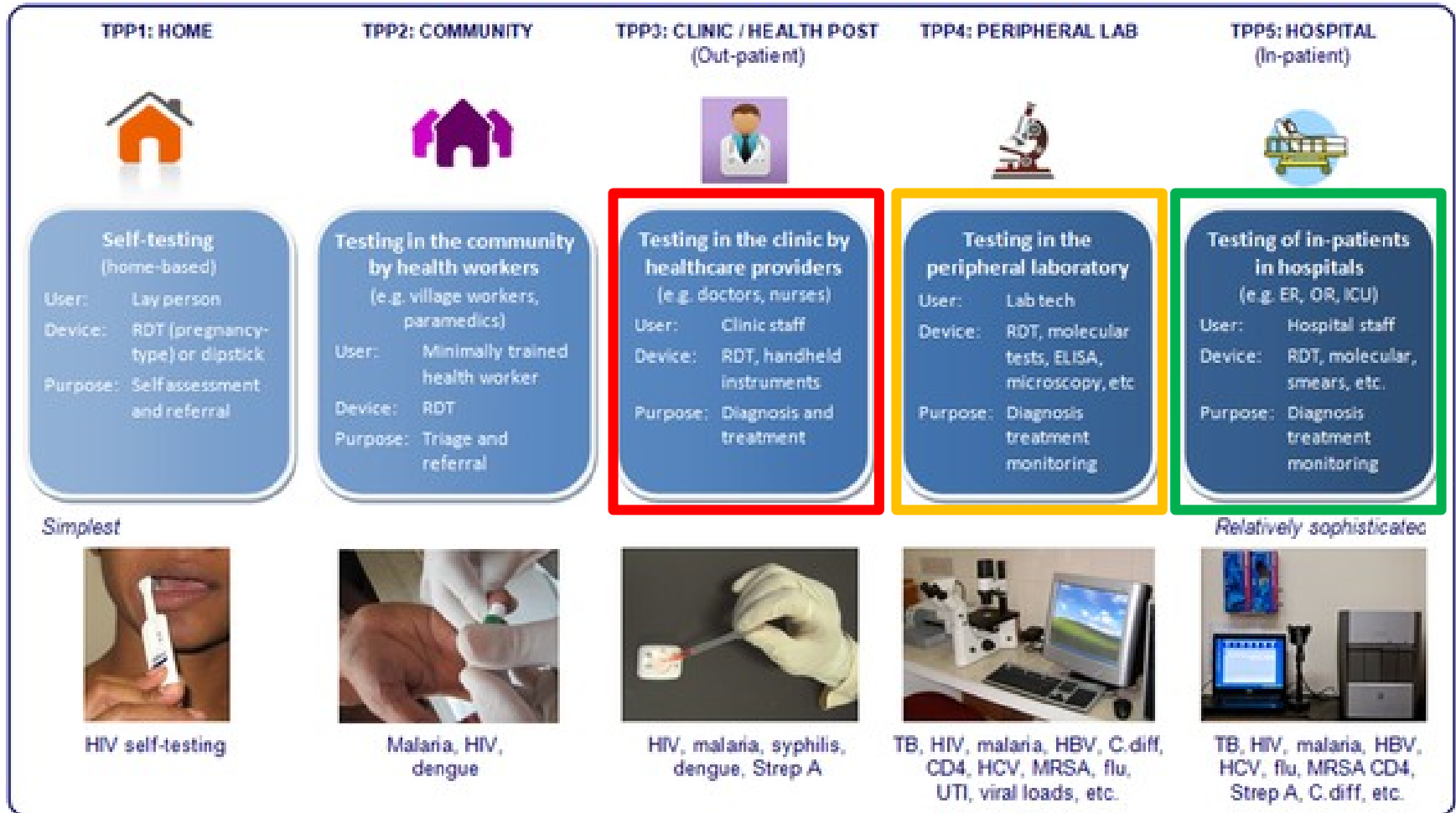
1 p.m. – 6 p.m.

Same day: species ID + MRSA / VRE / Carbapenemases

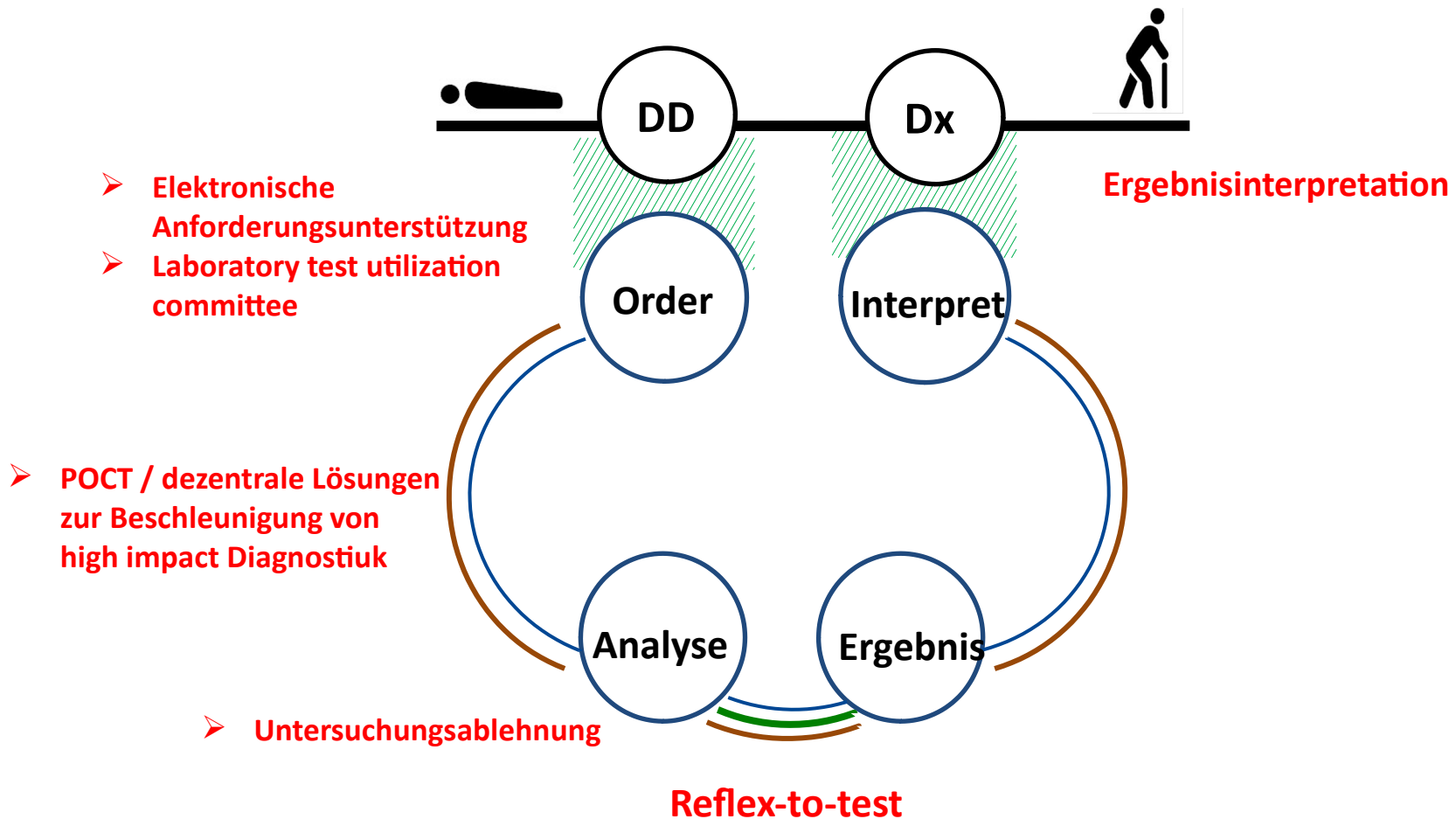
Assay	Pro	Con
Maldi-ToF ID	<ul style="list-style-type: none"> • Broad pathogen coverage • Relatively fast • Cheap • Widely available 	<ul style="list-style-type: none"> • No valid susceptibility prediction • Labor-intensive • Slow (4 h short incubation)
Multiplex molecular assays	<ul style="list-style-type: none"> • Fast • Short hands-on-time • Easy to use • Sensitive MRSA / VRE prediction 	<ul style="list-style-type: none"> • Limited pathogen coverage • Less sensitive in predicting Gram-negative resistance • Expensive
RAST (EUCAST)	<ul style="list-style-type: none"> • Cheap (consumables) • Reliable • Broad coverage of resistance mechanisms in Gram-negatives 	<ul style="list-style-type: none"> • Labor-intensive • Requires at least 4 h incubation time • Restricted to some species

Wie POC spPCR Assays zur Umstrukturierung diagnostischer Strukturen beitragen können





spPCR Analytik: sinnvoll wenn Ziele definiert und Assays strukturiert in Prozesse integriert werden



Kontakt

Prof. Holger Rohde
Institut für Medizinische Mikrobiologie, Virologie und Hygiene
Universitätsklinikum Hamburg-Eppendorf
Martinistrasse 52
D-20246 Hamburg

Email: rohde@uke.de